

# Genetic engineering of barley to improve *Fusarium* head blight resistance

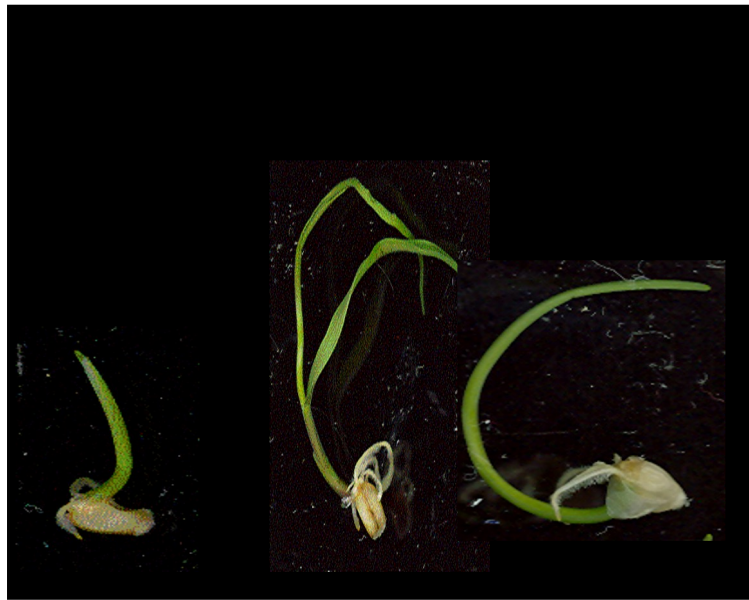
Rong Di

Rutgers University

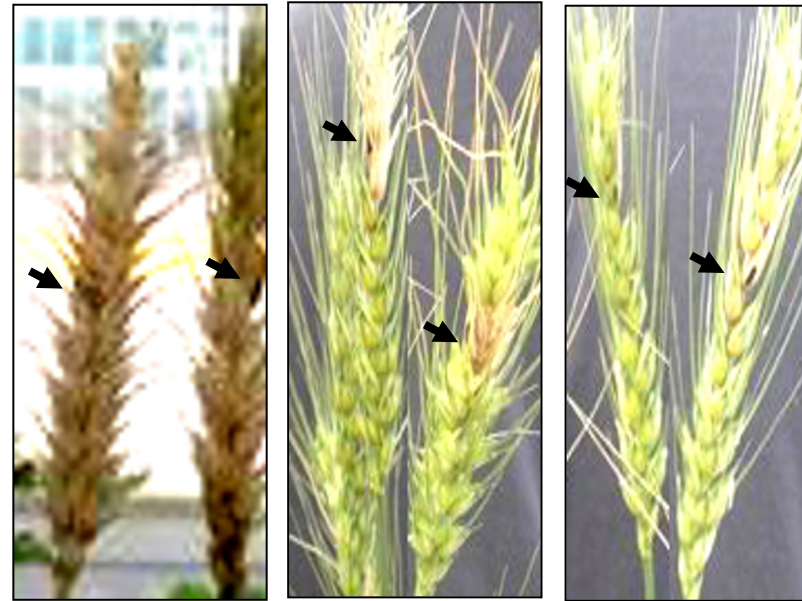


# Engineering FHB resistance in wheat and barley

Many transgenic genes have been engineered into mostly wheat to enhance FHB resistance including yeast ribosomal protein L3 (Di *et al.* , 2010) >>> **GMO**



**wt**      **RUT772**      **RUT8153**



**wt**      **RUT772**      **RUT8153**

- Di, R., A. Blechl, R. Dill-Macky, A. Tortora, and N. E. Tumer. 2010. *Plant Science* 178:374-380.
- U.S. Patent #8,026,410 B2. Tumer, N.E. and R. Di. Sept. 27, 2011. Transgenic plants expressing L3 delta proteins are resistant to trichothecene fungal toxins.

# CRISPR-gene editing to improve disease resistance

- **Knocking-out host disease susceptibility genes to interrupt plant-pathogen interaction**
- **gRNA and Cas9 transgene cassettes can be segregated from gene-mutated genomes.**
- **Transgene-free, gene-edited mutant plants can be produced in 1-2 generations.**

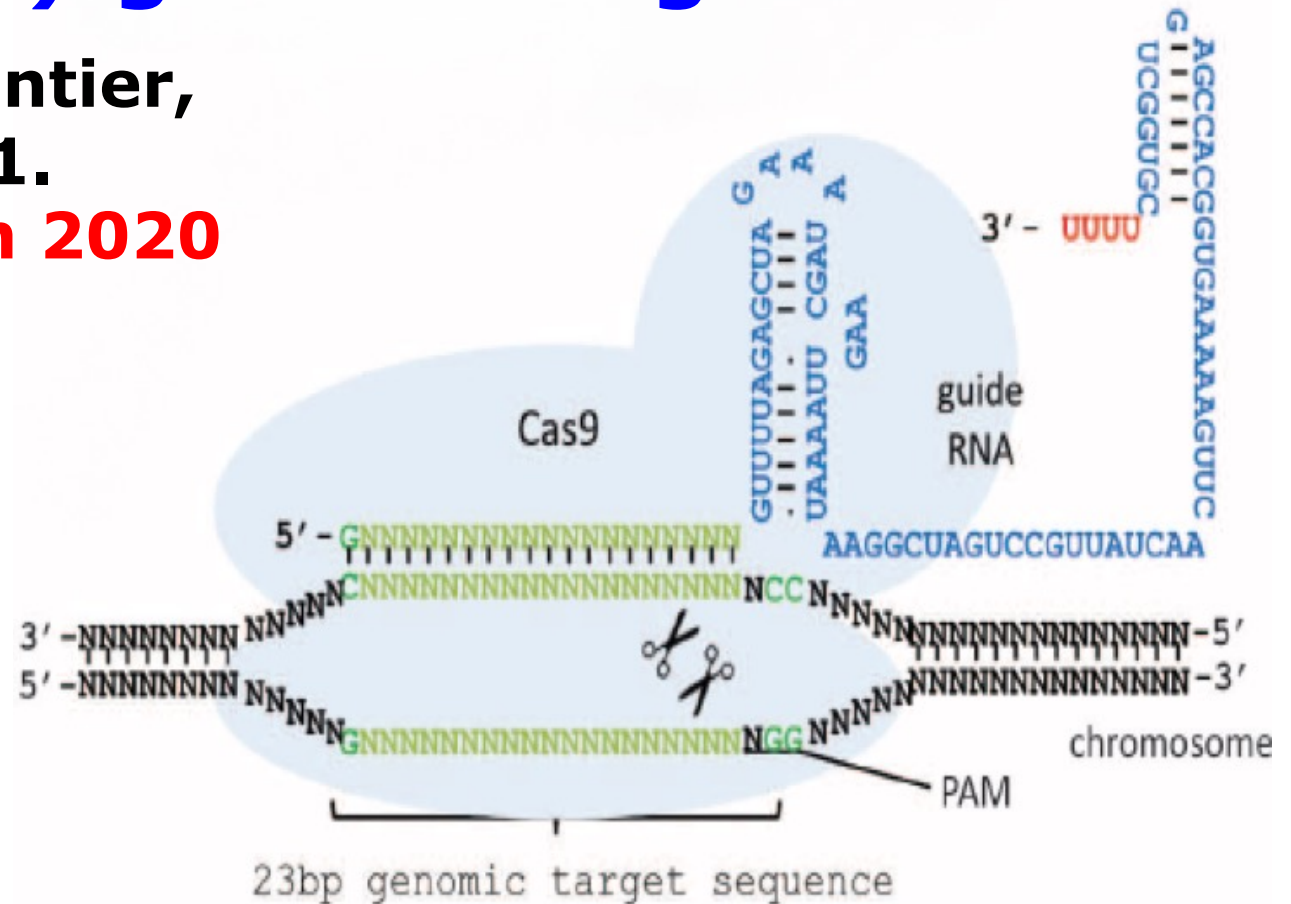
# CRISPR (Clustered Regularly Interspaced Short Palindromic Repeats)-gene editing

Jinek...Doudna and Charpentier,  
2012, Science 337:816-821.

**Nobel Prize in Chemistry in 2020**

## Key components:

- snRNA promoter
- gRNA (19-23 nt)
- scaffold
- Cas9 nuclease
  - codon optimized
- Double-stranded breaks (DSB) in gDNA

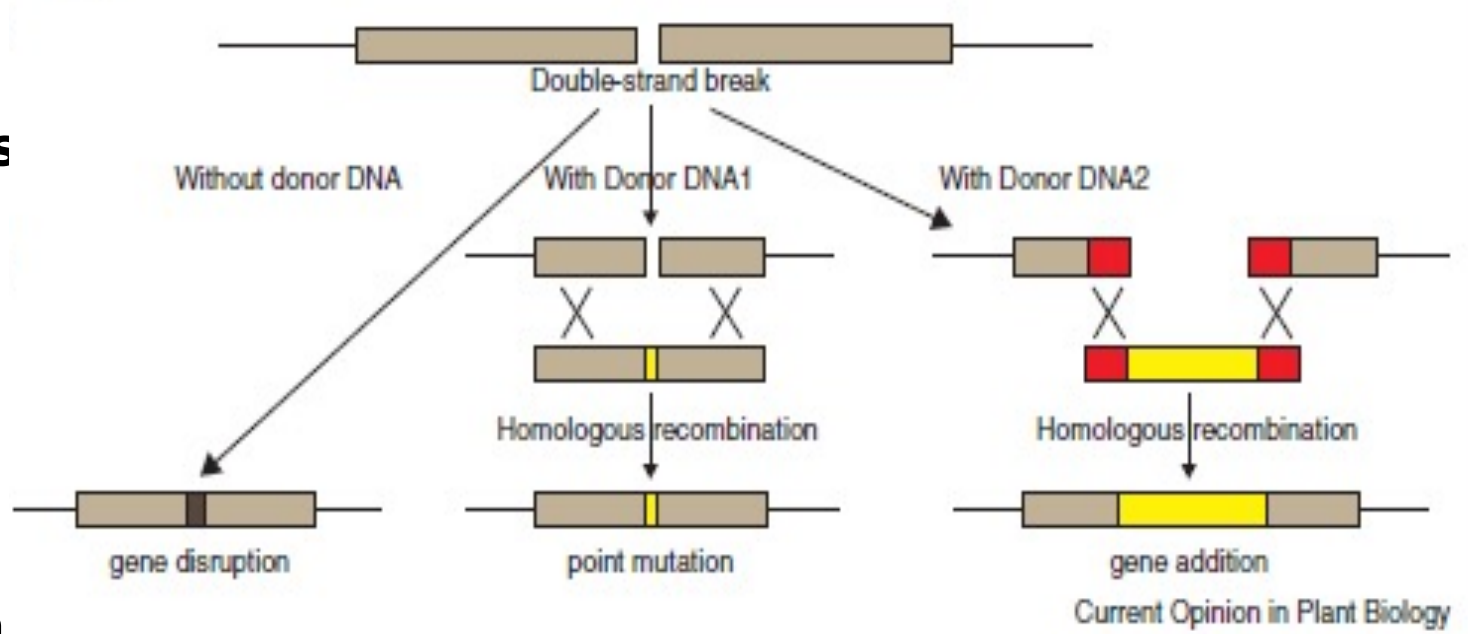


Mali et al., 2013, Science

# Mutations or gene editing by CRISPR

DSBs are repaired in nearly all cells by two highly conserved processes:

- **non-homologous end joining (NHEJ)**, which often results in small insertions or deletions (**Indels**), leading to gene disruption, gene editing.
- **homologous recombination (HR)**, which can be used for point mutation, gene insertion or replacement, gene editing.



Chen and Lin, 2013

# FHB in wheat and barley

## FHB in wheat

- *Fusarium* spp. infects wheat **during anthesis**
- colonizes florets first
- penetrates floral tissues
- spreads within spikes
- Two types of resistance:
  - Type I: resistance to initial infection
  - Type II: resistance to spread of infection

## FHB in barley

- *Fusarium* spp. infects **after anthesis**
- colonizes the brush hairs
- invades the developing caryopsis
- Natural Type II resistance;  
Symptoms do not spread in spikes



**Knocking out host factors to prevent initial infection will lead to effective FHB resistance in wheat and barley.**

(Huang, Muehlbauer et al. 2016)

# FHB susceptibility gene: 2OGO

## 2-oxoglutarate Fe(II)-dependent oxygenase (2OGO) gene

- Arabidopsis EMS-*At2OGO* mutant showed enhanced plant immunity to *Fg* infection. Knocking-out *At2OGO* gene seems to induce the expression of host defense genes. (Brewer *et al.*, 2014)
- *At2OGO* gene functions as salicylic acid 5-hydroxylase (S5H) to break down SA into gentisic acid and its impairment leads to SA accumulation. (Zhang *et al.*, 2017)
- We CRISPR-edited *At2OGO* specifically. *At2OGO*-KO plants are resistant to *F. graminearum*. Host defense genes are up-regulated.



- The *Hv2OGO*-complemented *At2OGO*-KO plants regained susceptibility to *Fg*, indicating *Hv2OGO* might function similarly as *At2OGO*.

Low, Y., M. A. Lawton and R. Di. 2020. Validation of barley 2OGO gene as a functional orthologue of Arabidopsis *DMR6* gene in *Fusarium* head blight susceptibility. *Sci. Reports*. 10:9935. DOI:10.1038/s41598-020-67006-5.

# FHB susceptibility gene: *EIN2*















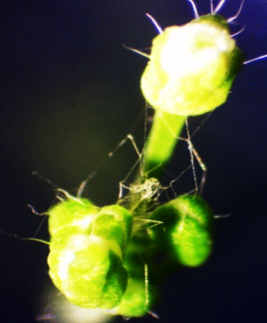



## **Ethylene insensitive 2 (*EIN2*) gene**

- ***Fg* exploits the ethylene signaling pathway via host *EIN2*-regulated signal transduction to promote susceptibility in both Arabidopsis and wheat. (Chen *et al.*, 2009)**
- **RNAi knocking-down of *EIN2* in wheat enhances FHB resistance. (Travella *et al.*, 2006)**
- **We CRISPR-edited *AtEIN2* specifically. *AtEIN2*-KO plants are resistant to *F. graminearum*. Ethylene signaling was down-regulated, but the SA and JA signaling pathways were unaffected.**
- **The *HvEIN2*-complemented *AtEIN2*-KO plants regained susceptibility to *Fg*, indicating *HvEIN2* might also function similarly as *AtEIN2*.**

Low, Y. C., M. A. Lawton and R. Di. 2022. *Ethylene insensitive 2 (*EIN2*) as a Potential target gene to enhance *Fusarium* head blight disease resistance. Plant Sci. 322:111361. DOI: 10.1016/j.plantsci.2022.111361.*



# The *AtEIN2*-KO plants were more resistant to *Fg* infection, compared to *At* WT and *AtEIN2*-KO/*HvEIN2*

Samples	Day 1	Day 2	Day 3	Day 4	Day 5	Day 6
<i>At</i> WT						
<i>AtEIN2</i> -KO						
<i>AtEIN2</i> -KO/ <i>HvEIN2</i>						

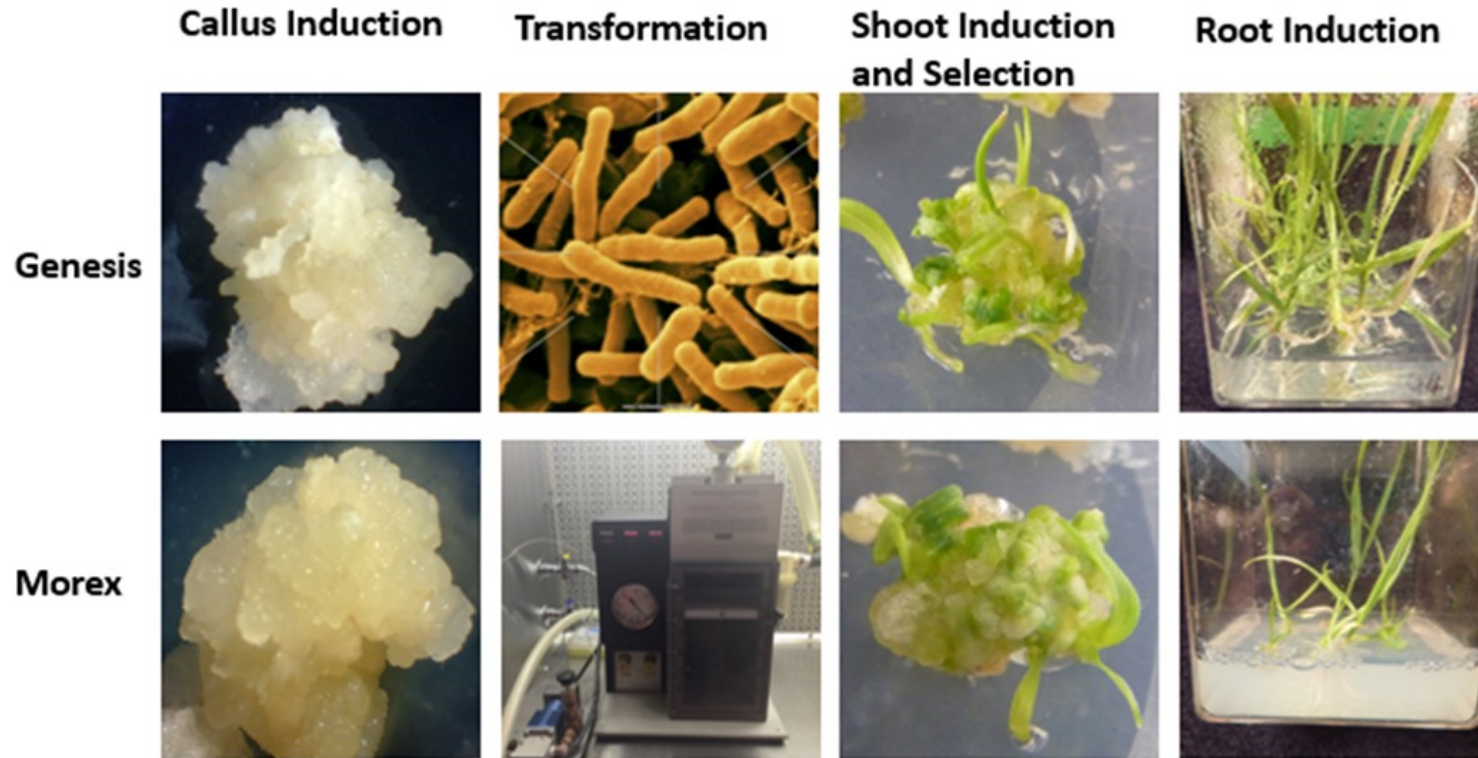
# Involvement of *HvUGT* gene in barley FHB resistance

Muehlbauer group (2016) showed that the *HvUGT* (*uridine diphosphate glycosyltransferase*, converting DON to the less toxic D3G ) is induced by DON in both the resistant and susceptible barley. So the *HvUGT* gene expression modulation is critical for FHB resistance in barley.

**Our barley FHB gene targets:**

***Hv20GO*, *HvEIN2* and *HvUGT* promoter**

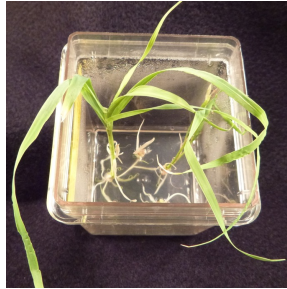
# Improved barley tissue culture and transformation system



- Embryogenic calli are induced from immature scutellum.
- Calli are transformed by biolistic bombardment or Agrobacterium.
- Multiple transgenic shoots are generated from calli initiated from a single immature seed.

# Production of barley *Hv20GO* mutant plants with pRD383 (POsU3/*Hv20GO*-gRNA::Cas9) by CRISPR-editing

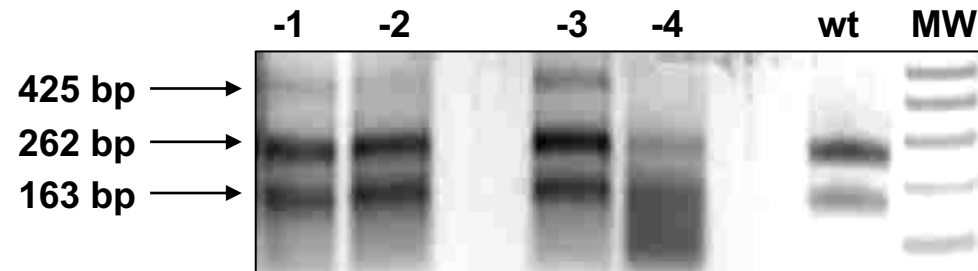
By Gene gun



By Agro



RFLP by *NdeI*



PCR-amplifying gDNA, cloning and sequencing

		<i>NdeI</i>									
WT	TCTACCCC	<u>AAG</u>	TGC	CCC	TCG	CCG	GAG	CTG	ACA	TAT	<u>GGGCTCCC</u>
383-C1	TCTACCCC	<u>AAG</u>	TGC	CCC	TCG	CCG	GAG	CA <u>G</u>	AC <u>G</u>	TAT	<u>GGGCTCCC</u>
								L204Q H205R			
383-C2	TCTACCCC	<u>AAG</u>	TGC	CCC	<u>CCG</u>	CCG	GAG	CTG	ACA	TAT	<u>GGGCTCCC</u>
					S201P						
383-CA2	TCTACCCC	<u>AAG</u>	TGC	CCC	<u>CCG</u>	CCG	GAG	CTG	ACA	TAT	<u>GGGCTCCC</u>
					S201P						

# Improvement of barley CRISPR-editing platform

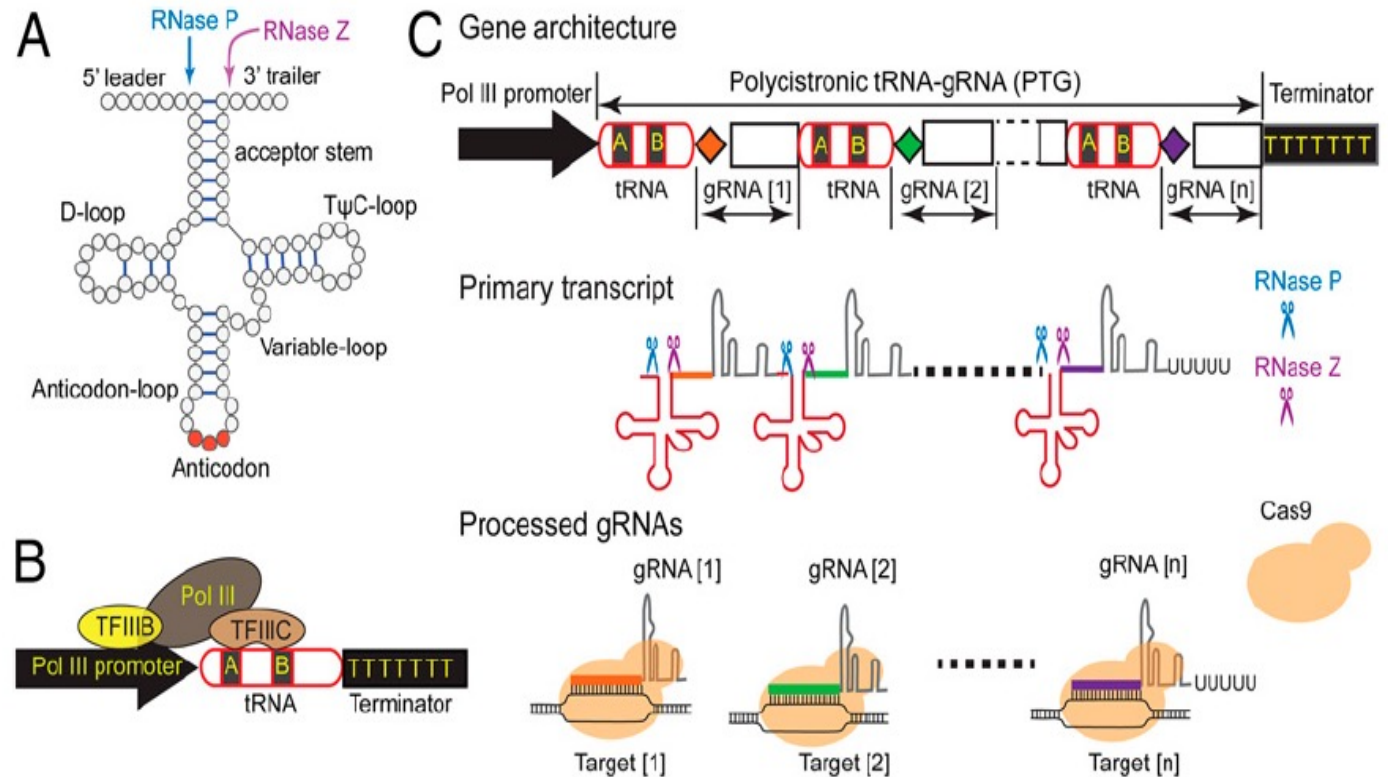
## Conventional plant CRISPR-gene editing platform

- $P_{OsU3}/Hv\ gRNA/P_{ZmUbi}::Cas9-Mo$ . The gRNA expression is not very efficient.

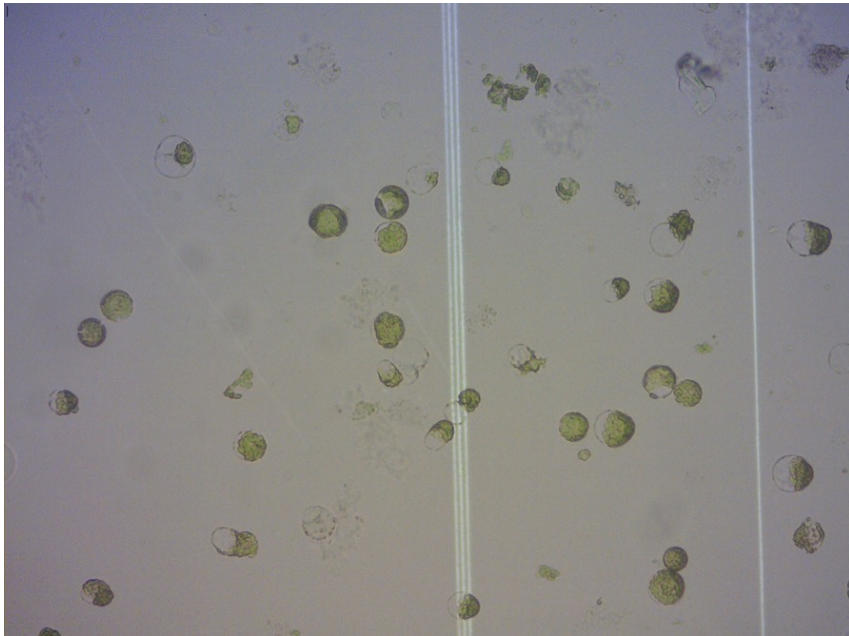
## The polycistronic tRNA-gRNA (PTG) platform

- Developed by the Yang group at Penn State U. (2015)
- tRNA processing is robust.
- No "G" or "A" requirement in the target sequence
- Multiplexing

**Dual barley CRISPR-editing vectors:**  
pRD543 (transient; dual *HvPUGT*)  
pRD549 (integrating; dual *HvPUGT*)  
pRD550 (transient; dual *HvEIN2*)  
pRD554 (integrating; dual *HvEIN2*)



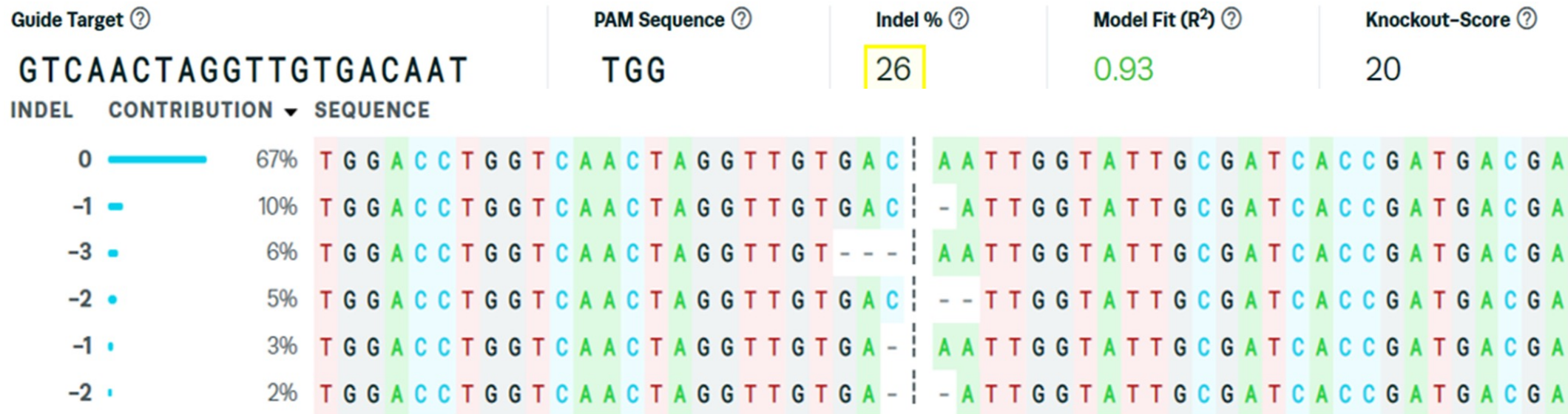
# Using barley protoplast to assess the mutagenesis efficiency of CRISPR-editing constructs



- **Isolate protoplast**
- **PEG-Ca<sup>2+</sup> mediated-transformation**
- **Isolate gDNA after 48 hr**
- **PCR-amplify gDNA spanning the target site**
- **Analyze for mutations**

# Using Synthego ICE CRISPR online tool to decipher the Indel frequency in the potential mutant tissues and plants

## *HvUGT* promoter target site

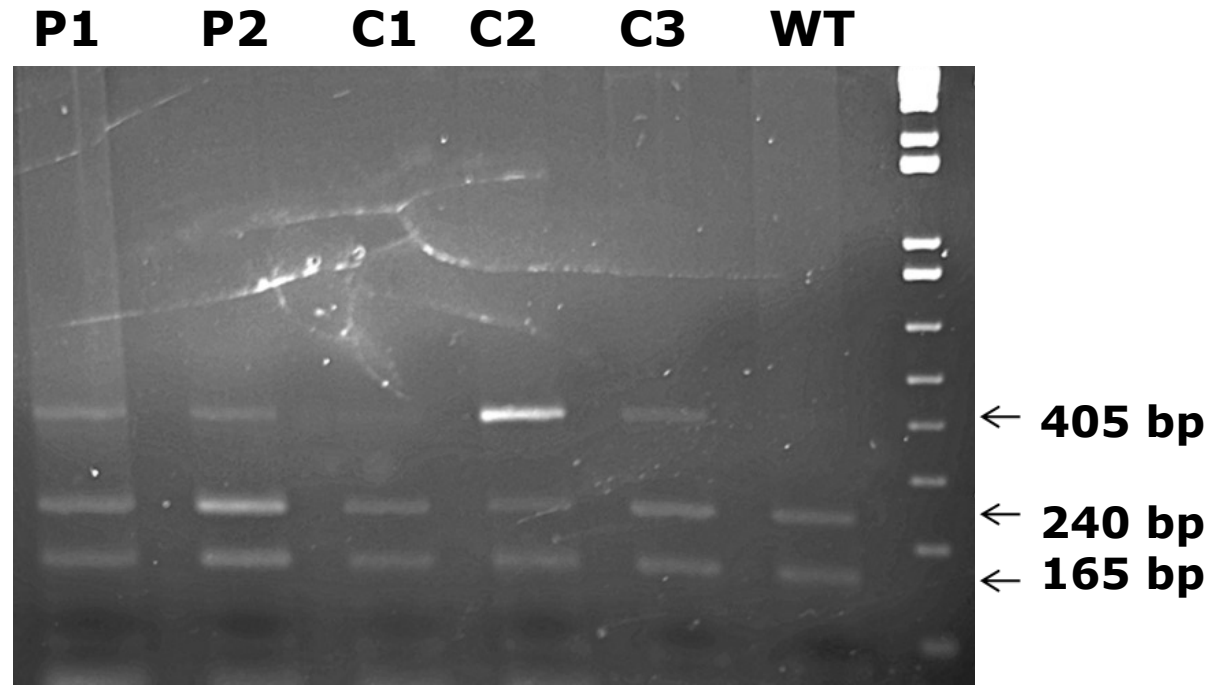
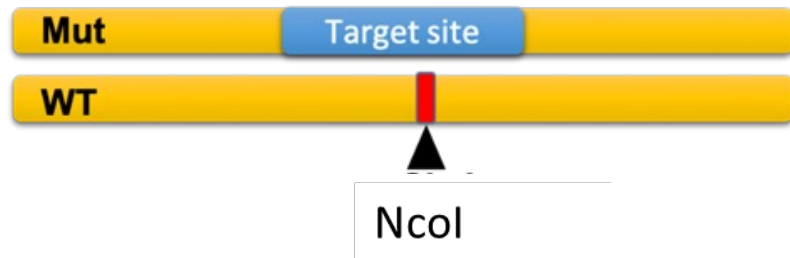


➤ Putative mutations were confirmed by Sanger sequencing of individual clones of PCR products.

# Restriction fragment length polymorphism (RFLP) assay to analyze CRISPR-induced mutation

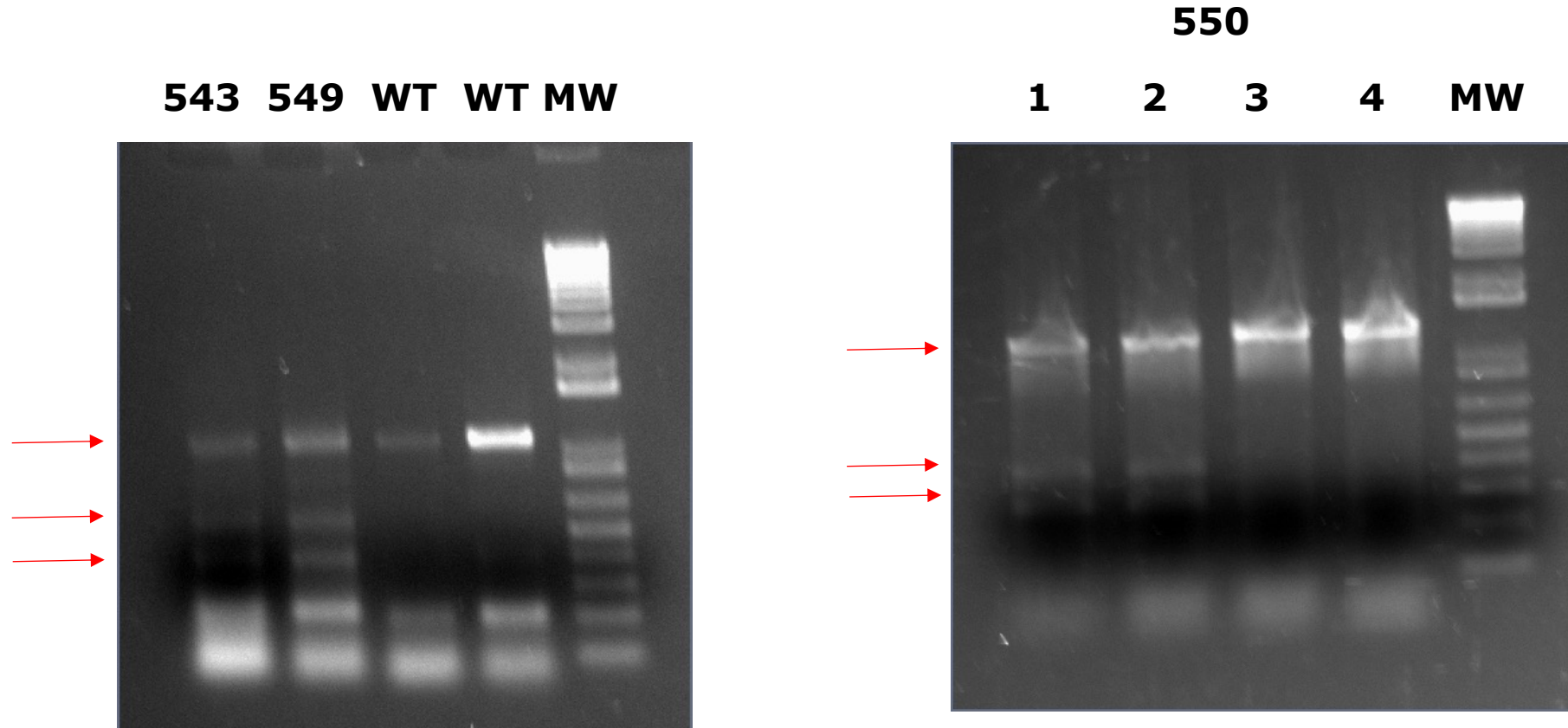
pRD549 (integrating, dual targeting *HvUGT* promoter)-transformed protoplasts (P) or callus (C)

PCR product of *HvUGT* promoter *Nco*I target site





# PCR-amplification and sequencing of gDNAs spanning the dual target sites of *HvGUT* promoter and *HvEIN2*



- The full-length products have mostly WT sequences.
- The shorter products have insertions from other barley gDNA parts.

# **Our gene-edited sweet basil plants produced by the transient CRISPR vector do not contain the transgene and are resistant to downy mildew disease**

**X. Zhang, Y. C. Low, M. A. Lawton, J. E. Simon and R. Di. 2021. CRISPR-editing of sweet basil (*Ocimum basilicum* L.) homoserine kinase gene for improved downy mildew disease resistance. *Frontiers in Genome Editing-Genome Editing in Plants* 3:629769. DOI:10.3389/fgeed.2021.629769.**

## Gene-edited CRISPR mushroom escapes US regulation

A fungus engineered with the CRISPR–Cas9 technique can be cultivated and sold without further oversight.

Emily Waltz

14 April 2016

# CHEMISTRY WORLD

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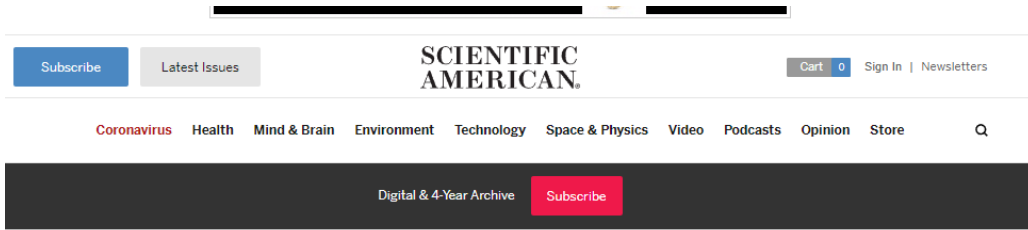
By Emma Stove, 26 April 2016

[Yinong Yang](#) and colleagues developed the anti-browning mushroom at Pennsylvania State University in the US. 'We used the Crispr-Cas9 technology to create small deletions in a specific gene encoding a polyphenol oxidase (PPO), says Yang.



The team used a bacterial plasmid construct to deliver the guide RNA and Cas9 enzyme into mushroom cells and achieve the necessary deletions. The final product does not contain any DNA from a donor or vector organism, so it is not covered by current regulations that control the import, movement or release of GMOs in the US.

2017



nature  
BIOLOGY  
**Genetically Modified Browning-Resistant Apple Reaches U.S. Stores**

Success for the “Arctic apple” could herald a new wave of lab-grown foods

By Amy Maxmen, Nature magazine on November 7, 2017

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### Availability of Deregulation Extension Request and Preliminary Determination for Apples Developed Using Genetic Engineering

Published: Jun 28, 2021

Print



**Okanagan Specialty Fruits, Inc. (OSF): non-browning apple**

-- PG451 Arctic Gala, GD743 Arctic Golden and GS784 Arctic Granny apple

-- RNAi silencing/Agrobacterium transformation to knock-down 4 PPO (polyphenol oxidase) genes to prevent browning



# Acknowledgement

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NJAES

**Collaborator:** Dr. M. Lawton

**Technician:** A. Dineen



# Q & A