

## Investigations of microbiological quality risks in nonalcoholic beer: growth potential of food-borne pathogens and alcohol formation by contaminant microbes

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Summary	Introduction
In this study, the growth potential of food-borne pathogens and unintended alcohol formation by contaminant microbes were investigated. We focused on the NAB <sup>*</sup> production process by dealcoholization, as well as the post-packaging distribution and on-premise sales of NAB kegs <sup>*</sup> . (*NAB sample: Alc.0.5 v/v%, pH4.6, 0.230 Mpa)	<ul> <li>Non-alcoholic beer (NAB)</li> <li>The definition of NAB differs from country to country. In many European countries, products are labeled as "non-alcoholic beer" when they contain less than Alc. 0.5 v/v%<sup>*1</sup>.</li> <li>NAB market is expanding. Furthermore, NAB keg have begun to be sold in recent years.</li> </ul>
<ul> <li>In NAB production by dealcoholization (Decarbonated NAB)</li> <li>✓ The growth potential of food-borne pathogens was low in NAB sample at 6°C.</li> <li>✓ Approximately 0.8% alcohol was formed by contaminant yeast during the period from the dealcoholization process to the completion of cooling. After cooling, the risk was controlled at 2°C in NAB sample.</li> <li>→ Process control and cold storage at around 2 °C is important after dealcoholization to prevent alcohol formation.</li> </ul>	<ul> <li>Microbial risks of NAB by dealcoholization</li> <li>Since NAB contains no or a very small amount of alcohol, it is considered that microorganisms grow more easily in NAB compared with traditional beer.</li> <li>The production process after dealcoholization is susceptible to microbial risks about the food-borne pathogens and alcohol formation, because of weaker gas pressure and alcohol.</li> <li>NAB keg is also more prone to microbial contamination after keg opening compared to</li> </ul>
To the next period distribution and an promise cales of NAD lasts	cans and bottles. There are concerns about the potential growth of food-borne pathogens

<ul> <li>In the post-packaging distribution and or</li> <li>✓ The growth potential of food-borne pathoge</li> <li>✓ The presence of preservatives tends to deaded</li> <li>✓ Alcohol formation was suppressed at low tere alcohol was formed by contaminant yeast at a supersence of sodium benzoate tends to supersence of the keg filling environe</li> <li>◆ Adding sodium benzoate were also considet tends to suppress the stored at low temperated tends to suppress tends tends tends to suppress tends tends to suppress tends te</li></ul>	ns was low in NAB. ctivate food-borne pathogens more rapidly. mperature for short period, although 0.5% or more t room temperature for long period. suppress the alcohol formation. ment is important to suppress alcohol formation. ered effective to control alcohol formation. sure to prevent alcohol formation after keg openin	<ul> <li>and unintended alcohol formation in on-premise consumption.</li> <li>In this study, We investigated the growth potential of food-borne pathogens and alcohol formation by contaminant microbes in commercial NAB produced by dealcolization (Alc.0.5 v/v%, pH4.6, 0.230 MPa). Regarding the growth potential of food-borne pathogens during the cold storage, we focused on <i>E. coli</i> and <i>Salmonella sp.</i>, because they are considered to have stronger growth potential in NAB *2. Our goal is to find effective control measures against food-borne pathogens and alcohol-forming contaminants (below 1% ABV).</li> </ul>		
	Results and	Discussion		
1. The microbial ris by dealc	ks in NAB production oholization	2. The microbial risk and on-p	ks in the post-packagin remise sales of NAB ke	g distribution gs
1-1. Control measures of food- after dealcoholization pro	borne pathogens cess	<b>2-1. The growth potenti</b> <u>Materials and Methods</u> : <i>• E. coli</i> 0157	al of food-borne patho A) E. coli 0157	<b>gens</b> <b>B)</b> Salmonella enterica 10 <sup>6</sup>
Materials and Methods:Strain• E. coli NBRC3301• Salmonella enterica sub sp. enterica NBRC12529SampleDecarbonated NAB (pH4.6)	[6°C, 100 days]	CRA16244 CRA16039 CRA16040 • Salmonella enterica CRA1868 CRA1947	10 <sup>4</sup> 10 <sup>2</sup>	10 <sup>4</sup> Potassium pyrosulfit
Condition* • 1000 cells/ml • 6°C, 100 days • Aerobic incubation without agitation • n=2	$\begin{bmatrix} 10^{2} \\ 10^{1} \\ 10^{1} \\ 10^{0} \end{bmatrix}$ $\begin{bmatrix} 10^{2} \\ 10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <10 \\ <1$	Strain CRA3736 <i>Strain</i> <i>S. aureus</i> CRA2095 CRA1208 CRA11018	10 <sup>0</sup> 0 30 60 90 <b>C)</b> <i>S. aureus</i> 10 <sup>6</sup>	10 <sup>0</sup> 0 30 60 90 <b>D) Yersinia enterocolitica</b> 10 <sup>6</sup>
<ul> <li>★ 6°C, 100 days: After cooling process</li> <li><u>Results</u>:</li> <li>➢ No growth was observed with tested food-</li> </ul>	0 20 40 60 80 100 day Fig. 1. The growth potential of	<ul> <li>Yersinia enterocolitica</li> <li>CRA4103</li> <li>CRA498</li> <li>CRA499</li> </ul>	$\overline{E}$ 10 <sup>4</sup>	104

 No growth was observed with tested foodborne pathogens at 6°C for 100 days (Fig.1).
 Salmonella enterica decreased more slowly than *E. coli* in NAB (Fig.1). Fig. 1. The growth potential of *E. coli* and *Salmonella enterica* at 6 °C (n=2)

## **1-2. Control of alcohol-forming contaminants**

Materials	and Methods:	Table 1. Tested strains
Strain	Yeast and bacteria (Table 1)	Tested strains
Sample	Decarbonated NAB (pH4.6)	(1) Brewing yeast (bottom) (2) Saccharomyces cerevisiae (S. cerevisiae) NBRC565
Condition*	<ul> <li>1000 cells/ml</li> <li>15°C, 5 days</li> <li>2 or 4°C, 100 days</li> <li>Anaerobic incubation</li> <li>n=3</li> </ul>	<ul> <li>(3) S. cerevisiae NCYC1236</li> <li>(4) S. cerevisiae var. diastaticus IFO1440</li> <li>(5) S. cerevisiae var. diastaticus ATCC13007<sup>T</sup></li> <li>(6) S. bayanus NBRC11022</li> <li>(7) Brettanomyces naardenensis (B. naardenensis) AGYC109</li> <li>(8) Candida parancilogis (C. parancilogis) ACYC41</li> </ul>
<ul> <li>*15°C, 5 da</li> <li>: During the process to the 2 or 4°C, 1</li> <li>: After coolin</li> </ul>	<b>ys</b> period from the dealcoholization e completion of cooling <b>.00 days</b> g process	<ul> <li>(0) Canadida parapsilosis (C. parapsilosis) AGTC41</li> <li>(9) C. parapsilosis AGYC176</li> <li>(10) Zygosaccharomyces rouxii (Z. rouxii) NBRC1130<sup>T</sup></li> <li>(11) Z. rouxii IFO0495</li> <li>(12) Z. fermentati NBRC0479<sup>T</sup></li> <li>(13) Z. bailii AGYC120</li> <li>(14) Levilactobacillus brevis (Lactobacillus brevis) JCM1170</li> <li>(15) Levilactobacillus brevis ABBC64</li> <li>(16) Secundilactobacillus paracollinoides (Lactobacillus paracollinoides) JCM11969<sup>T</sup></li> <li>(17) Fructilactobacillus lindneri (Lactobacillus lindneri) DSM20690<sup>T</sup></li> <li>(18) Pediococcus damnosus ABBC478</li> </ul>
1.0 —	[15°C, 5 days]	1.0 [2 or 4°C, 100 days]
0.8		0.8
(%) 0.6 ⊲ylc 0.4		0.6 2 °C 0.4 0.4 0.4





of food-borne pathogens at 25 °C (n=2)

No growth was observed with food-borne pathogens, independent of preservative addition (Fig.4).

The presence of preservatives tends to deactivate *E. coli* O157, *Salmonella enterica*, and *Yersinia enterocolitica* more rapidly in NAB samples (Fig.4).

## **2-2. Alcohol formation and control of contaminant microbes**

Materials and Methods:

Results

 Table 2. Tested strains

Strain	Yeast and bacteria (Table 2)		Tested strains	
	Carbonated NAB (pH4.6)	Decarbonated NAB (pH4.6)	<ul> <li>(1) Brewing yeast (bottom)</li> <li>(2) S. cerevisiae NBRC565</li> <li>(3) S. cerevisiae var. diastaticus IFO1440</li> </ul>	
Sample	<ul> <li>Potassium pyrosulfite (40 ppm)</li> </ul>		(4) S. cerevisiae var. diastaticus $ATCC13007^{T}$ (5) S. bayanus NBRC11022 (6) B. naardenensis AGYC109 (7) Dekkera anomala (D. anomala) $ATCC10562^{T}$	
Condition	<ul> <li>1000 cells/ml</li> <li>25°C, 3 months</li> <li>n=2</li> </ul>	<ul> <li>1000 cells/ml</li> <li>4°C, 1 week</li> <li>4°C, 2 weeks</li> <li>Anaerobic incubation</li> <li>n=2</li> </ul>	<ul> <li>(8) <i>D. anomala</i> NBR</li> <li>(9) <i>D. anomala</i> ATCO</li> <li>(10) <i>Z. fermentati</i> NE</li> <li>(11) <i>Z. bailii</i> AGYC12</li> <li>(12) <i>Pediococcus dari</i></li> </ul>	C0642 C10559 BRC0479 <sup>T</sup> 0 <i>nnosus</i> ABBC478
A) 25°C, 3 months     B) 4°C, 1       1.0     1.0		L week 1.0	C) 4°C, 2 weeks	
0.8	0.8		0.8	Control Sodium benzoate
0.6 —	0.6		0.6	Potassium pyrosulfite



- In NAB production; The growth of Salmonella enterica and E. coli were controlled at 6°C or less after dealcoholization. Additionally, alcohol was formed by contaminant yeast during the period from the dealcoholization process to the completion of cooling. After cooling, alcohol formation was suppressed at 2°C, although approximately 0.8% alcohol was formed at 4°C.
- In the NAB kegs; The growth potential of food-borne pathogens was controlled. Alcohol formation by contaminant yeast was controlled at low temperature for short period. Although 0.5% or more alcohol was formed at room temperature for longer period, the alcohol formation was suppressed by adding sodium benzoate (400 ppm).

→ Controlling the production process with storage temperature at around 2°C after dealcoholization is important to suppress alcohol formation. Additionally, the microbiological management of the NAB keg filling environment is critical to suppress alcohol formation. If there are concerns about the filling environment, adding sodium benzoate is considered effective. Furthermore, NAB kegs should be stored at low temperature to prevent unintended alcohol formation after keg opening.

**References** 1. Nils Rettberg *et.al. Beverages*, **8** (1), 4 (2022). 2. Garry Menz *et.al. J. Food.Prot.*, **74** (10): 1670–1675 (2011).